# New pharmacological strategies for treatment of Alzheimer's disease: focus on disease modifying drugs

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Current approved drug treatments for Alzheimer disease (AD) include cholinesterase inhibitors (donepezil, rivastigmine, galantamine) and the NMDA receptor antagonist memantine. These drugs provide symptomatic relief but poorly affect the progression of the disease. Drug discovery has been directed, in the last 10 years, to develop 'disease modifying drugs' hopefully able to counteract the progression of AD. Because in a chronic, slow progressing pathological process, such as AD, an early start of treatment enhances the chance of success, it is crucial to have biomarkers for early detection of AD-related brain dysfunction, usable before clinical onset. Reliable early biomarkers need therefore to be prospectively tested for predictive accuracy, with specific cut off values validated in clinical practice. Disease modifying drugs developed so far include drugs to reduce  $\beta$  amyloid ( $A\beta$ ) production, drugs to prevent  $A\beta$  aggregation, drugs to promote  $A\beta$  clearance, drugs targeting tau phosphorylation and assembly and other approaches. Unfortunately none of these drugs has demonstrated efficacy in phase 3 studies. The failure of clinical trials with disease modifying drugs raises a number of questions, spanning from methodological flaws to fundamental understanding of AD pathophysiology and biology. Recently, new diagnostic criteria applicable to presymptomatic stages of AD have been published. These new criteria may impact on drug development, such that future trials on disease modifying drugs will include populations susceptible to AD, before clinical onset. Specific problems with completed trials and hopes with ongoing trials are discussed in this review.

#### Introduction

Alzheimer's disease (AD) is a common disorder characterized by cognitive decline [1] associated with the presence of  $\beta$ -amyloid (A $\beta$ ) in plaques, intracellular aggregates of tau protein, forming neurofibrillary tangles (NFT) and progressive neuronal loss [2]. A $\beta$  plays a primary role in AD pathophysiology [2]. Oligomer species of aggregated A $\beta$  exert toxic effects on synaptic and cellular functions, finally leading to neurodegeneration and cognitive, as well as neuropsychiatric, symptoms [3]. Current treatment of AD includes cholinesterase inhibitors (donepezil, rivastigmine, galantamine), used for mild to moderate AD, and the NMDA receptor antagonist, memantine, approved for the treatment of moderate to severe AD [4, 5]. These drugs mainly provide symptomatic, short-term benefits, without

affecting the underlying pathogenic mechanisms of the disease [4], though a neuroprotective potential has also been proposed [6, 7]. Developing disease modifying drugs, able to counteract the progression of AD, is one of the biggest challenges of modern pharmacology. The pathophysiological process of AD begins many years before clinical diagnosis is set. The optimal time for disease-modifying drug treatment may therefore be in the presymptomatic stage of AD, where the disease is still hidden. Recently, the criteria for the clinical diagnosis of AD have been revised by the National Institute on Ageing and the Alzheimer's Association workgroup [8]. The new criteria incorporate biomarkers to identify early stages of AD, susceptible to being treated with disease modifying drugs [9, 10].

In the present review, we will summarize the new pharmacological strategies for the treatment of AD, focusing



 Table 1

 Current status of clinical development of some disease modifying drugs for treatment of Alzheimer's disease (AD)

Drug	Mechanism of action relevant for AD	Phase of study	Result of study	Caveat of study
Rosiglitazone	β-secretase inhibition (?)	3	Ineffective	Lack of biomarker
Semagacestat	γ-secretase inhibition	3	Premature end	Severe adverse drug reaction
Tarenflurbil	γ-secretase modulation	3	Ineffective	Low potency, blood-brain barrier passage
Tramiprosate	Inhibition of Aβ oligomerization	3	Ineffective	-
Scyllo-inositol	Inhibition of Aβ oligomerization	2	Ineffective	Biomarker change
Bapineuzumab	Aβ clearance	3	Ongoing	Vasogenic oedema, amyloid angiopathy
Solaneuzumab	Aβ clearance	3	Ongoing	-
Lithium	Inhibition of tau phosphorylation	2	Clinical improvement Decrease of P-tau in CSF	-
Methylthioninium chloride	Inhibition of tau aggregation	2	Clinical improvement with 60 mg day <sup>-1</sup>	Lack of biomarker
Nilvadipine	Aβ clearance	Open label	Clinical improvement	Lack of biomarker
Latrepirdine	Mitochondrial protection	3	Ineffective Ongoing (in association with other drugs)	-

our attention on potential disease modifying drugs currently studied in phase 3 clinical trials. A summary of the current status of the clinical development of some disease modifying drugs is shown in Table 1.

## Disease modifying drugs: definition and implications for drug development in AD

A disease modifying drug is an agent that slows the progression of structural damage, such that its effect is persistent and can be detected even after stopping the treatment, because the cumulative pathological changes would be less severe in the treated group as compared with the control (placebo) group. In contrast, the definition 'symptomatic drug' refers to an agent that does not alter the progression of the disease, but only decreases (palliates) the severity of symptoms. The symptomatic effect is usually reversible, such that, if the treatment is interrupted, the treated group might be indistinguishable from the control (placebo) group. Definition and validation of appropriate biomarkers and scales of clinical outcome are of paramount importance for assessing efficacy of disease modifying drug treatments for AD. Agents that target the underlying pathophysiology of AD are expected to have greater effect on biomarker levels and disease progression before any substantial, irreversible functional loss occurs [11]. Biological markers of AD may be divided into different classes according to the 'amyloid' hypothesis. Biomarkers of brain A $\beta$  amyloidosis include both reduction in A $\beta_{42}$  in cerebrospinal fluid (CSF) [12] and positron-emission tomography (PET) evidence of AB deposition, using a variety of specific ligands [13]. Elevated tau in CSF seems related to neuronal injury, but is not specific for AD. However, the association of elevated tau with low concentrations of  $A\beta_{42}$  in CSF is considered the most informative

biomarker of AD. Furthermore, low  $A\beta_{42}$  in CSF together with elevated tau might help in predicting the progression of patients with mild cognitive impairment (MCI) to AD [9]. In this respect, a recent report shows, in a presymptomatic carrier of an APP mutation, decrease of  $A\beta_{42}$  and increase tau concentrations in CSF, with substantial changes in a 5 year, symptom free, interval [14]. Further studies are needed, both in early onset AD and late onset AD patients, to confirm whether these CSF biomarkers might be sensitive indicators of presymptomatic disease.

Other biomarkers, such as PET measurement of fluorodeoxyglucose 18F (FDG) uptake and magnetic resonance imaging (MRI) of brain atrophy, track indices of synaptic dysfunction and neuronal injury and are less specific [12]. However, all together these biomarkers may be very helpful in the early detection of AD-related brain dysfunction. In fact, studies conducted in carriers of AD genetic risk factors, have demonstrated the presence of AB accumulation in CSF, positive PET amyloid imaging, FDG-PET hypometabolism and functional MRI abnormalities up to a decade before the clinical onset of AD [10, 12]. These biomarkers need to be prospectively tested for predictive accuracy. Moreover, specific cutoff values need further validation in clinical practice. Neuropsychological and neurobehavioural tools to detect the earliest clinical manifestations of AD might be particularly useful in monitoring the response to disease modifying therapies in amnestic MCI patients, that have a prominent impairment in episodic memory and positive biomarkers [9]. Because AD is slowly progressing, demonstrating the effectiveness of a disease modifying treatment might require years. Most clinical studies examine 18-24 months of active treatment compared with placebo, but should provide informative data for a much longer period of time, given that patients are likely to take these medications for many years in clinical practice.

Up to now no disease modifying drugs are available for AD. Several have been tested, down to phase 3, but none

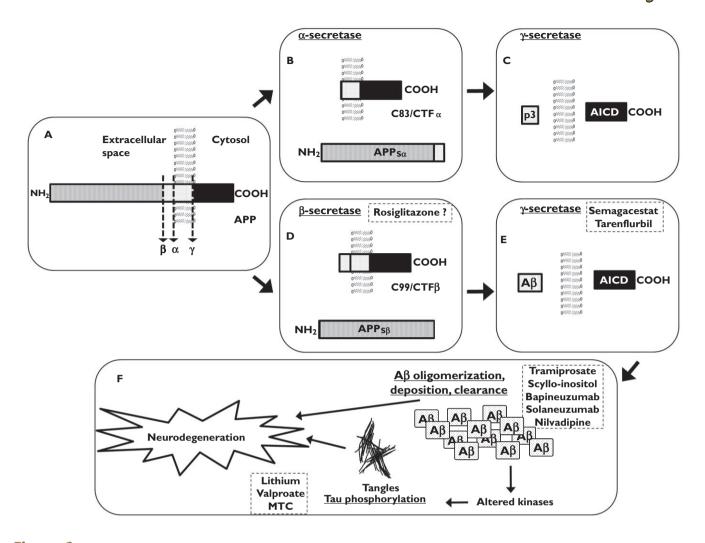
has yet reached approval. The failure of clinical trials with disease modifying drugs raises a number of guestions, spanning from methodological flaws to fundamental understanding of AD pathophysiology and biology. Some problems may arise from publication bias that favours positive results [15, 16], biomarkers and clinical outcomes utilized in animal models that substantially differ from human studies and time course of treatment in relation to development of disease, i.e. clinical studies enrol symptomatic patients, where some degree of neurodegeneration is already in place. Since the original Alzheimer's description [17], AB production and deposition has been considered as the main activity responsible for the pathological mechanism of AD, because it was documented in amyloid plaques of AD subjects by post mortem analysis. This view is referred to as the 'A $\beta$  hypothesis'. The A $\beta$ hypothesis has recently been challenged by the observation that  $A\beta$  clearing is not necessarily accompanied by cognitive improvement [2, 18, 19]. The physiological role of Aβ peptides, encoded also in the genome of the normal (healthy) population, has just begun to be unravelled and might be involved in basic mechanisms of cognition and memory, such as long-term potentiation (LTP) [20]. Proper folding and aggregation state of Aβ, rather than its absolute concentration, seems to be the determinant of neuronal toxicity in AD [21]. While assessing Aβ folding and aggregation state in vitro or post mortem in brain tissues is achievable [22, 23], this is not feasible, at present, in the living human brain, which makes the use of parenchymal Aβ as an AD biomarker very difficult.

#### Drugs to reduce Aβ production

As shown in Figure 1, generation of  $A\beta_{40}$  or  $A\beta_{42}$  is the result of two sequential cleavages of the amyloid precursor protein (APP). First, extracellular cleavage of APP by  $\beta$ -secretase 1 (also termed beta-site amyloid precursor protein cleaving enzyme 1 or BACE1) produces a soluble extracellular fragment and a cell membrane-bound fragment referred to as C99. Subsequent cleavage of C99 within its transmembrane domain by  $\gamma$ -secretase releases the intracellular domain of APP and generates A $\beta$  (Figure 1). In contrast, initial cleavage of APP by  $\alpha$ -secretase prevents generation of A $\beta$ , because, by cleaving APP closer to the cell membrane than  $\beta$ -secretase does, it removes a fragment of A $\beta$  (Figure 1) [24]. Therapeutic attempts have targeted inhibition of  $\beta$ -secretase and  $\gamma$ -secretase.

 $\beta$ -secretase 1 is an aspartyl protease that shares some features with HIV aspartyl proteases [25]. No known mutations in the gene encoding  $\beta$ -secretase have been related to familial AD, but elevated levels of this enzyme have been found in sporadic AD [26] and might be associated with polymorphism in the promoter region [27]. Because  $\beta$ -secretase 1 also has other substrates (including neuregulin-1, which is involved in myelination), develop-

ment of inhibitors may theoretically face problems of toxicity related to non-specific effects, though deletion of the β-secretase 1 gene produces only minor phenotype changes [28]. The thiazolidinediones, rosiglitazone and pioglitazone, that have been tested for AD in randomized controlled trials (RCTs), may in part act as suppressors of β-secretase expression [29]. Chang et al. reported recently [30] that the administration of a  $\beta$ -secretase inhibitor rescued cognitive decline and reduced brain AB in AD mice Tg2576, with no toxicity over a 7 month time period. Up to now no efficacy data are available from phase 3 clinical trials of  $\beta$ -secretase inhibitors. Specific problems in developing safe, non-toxic β-secretase inhibitors are related to blood-brain barrier (BBB) penetration and reasonable selectivity. Some interesting compounds have been designed by using crystal structure based inhibitor design [31] and some have been tested or currently are in phase 1 trials [32]. As mentioned above, rosiglitazone is an antidiabetic drug that has been clinically tested in AD. The main mechanism of action of rosiglitazone in diabetes, i.e. PPARy binding and subsequent transcription of genes involved in metabolic control, is precisely defined at the molecular level [33]. The same, however, cannot be stated for a supposed beneficial effect of rosiglitazone in AD. Starting from a correlation between insulin resistance and AD [34], preclinical studies looked for an effect of rosiglitazone in animal models of AD, without, however, a precisely defined testable hypothesis in terms of molecular and cellular mechanisms [35]. Rosiglitazone was shown to improve spatial learning and memory abilities, slightly decrease  $A\beta_{42}$  concentrations in brain (but not  $A\beta_{40}$ ) and induce insulin-degrading enzyme (IDE), without affecting the amyloid plaque burden in Tg2576 mice [36]. IDE is a thiol metalloprotease that degrades insulin as well as monomeric Aβ [37], whose expression has been shown to be PPARy-dependent in neurons [38]. However, the quantitative contribution that IDE may give to Aβ turnover in brain parenchyma remains to be determined, and PPAR $\delta$ , rather than PPARy, may have a stronger effect in expression of Aβ degrading enzymes [39]. In one phase 2 study, after 6 months treatment with rosiglitazone, patients with mild AD or amnestic MCI exhibited better delayed recall and selective attention as compared with the placebo group [40]. The only biomarker used was A $\beta$  in plasma, which was decreased in the placebo group [40]. This finding was interpreted by the authors as an index of A $\beta$  deposition in brain, potentially contributing to clinical worsening, an explanation that cannot be considered satisfactory, because, at variance with A $\beta$  in CSF [13, 41], circulating A $\beta$  does not provide reproducible correlation with AD [42] and does not reflect A $\beta$  processing in the brain [43]. In another phase 2 study, mild to moderate AD patients were treated with three different doses of rosiglitazone for 24 weeks and the data were stratified according to the APOE  $\varepsilon 4$  allele status. In APOE ε4 non-carriers rosiglitazone seemed to determine a cognitive and functional improvement, whereas APOE £4



#### Figure 1

Main steps of sequential cleavage of amyloid precursor protein (APP), leading to generation of  $\beta$  amyloid (A $\beta$ ) and/or other products. In A, dashed arrows indicate the cleavage sites for  $\alpha$ -,  $\beta$ - and  $\gamma$ -secretase. In B, ectodomain shedding of APP by  $\alpha$ -secretase gives a soluble extracellular APP fragment (APPs $\alpha$ ) and a 83 amino acid, membrane-bound, carboxy terminus fragment (C83/CTF $\alpha$ ). Subsequent intramembrane proteolytic cleavage of C83/CTF $\alpha$  by  $\gamma$ -secretase (in C) releases a short extracellular p3-peptide (p3) and a cytosolic APP intracellular domain (AICD). In D, ectodomain shedding of APP by  $\beta$ -secretase gives a soluble extracellular APP fragment (APPs $\beta$ ) and a 99 amino acid, membrane-bound, carboxy terminus fragment (C99/CTF $\beta$ ). Subsequent intramembrane proteolytic cleavage of C99/CTF $\beta$  by  $\gamma$ -secretase (in E) releases extracellular A $\beta$  and a cytosolic APP intracellular domain (AICD). In F, A $\beta$  oligomerization and deposition lead to neurodegeneration, both directly and through tau hyperphosphorylation. In dashed boxes, disease modifying drugs that interfere with each particular step. See text for more information and references

allele carriers showed no beneficial effect [44]. Of note, no specific AD biomarker was assessed in this study. Analysis of data according to the APOE  $\epsilon$ 4 allele status was based on the hypothesis that  $\epsilon$ 4-positive individuals, susceptible to early onset AD [45], have disturbances in enzymatic pathways of glucose metabolism in the brain, reminiscent of insulin resistance as seen in type 2 diabetes [46, 47], that might be sensitive to PPAR $\gamma$  activation by rosiglitazone. In contrast with the initial hypothesis, however, a mild clinical benefit was seen in  $\epsilon$ 4 non-carriers. The subsequent larger phase 3 study showed no significant clinical benefit of rosiglitazone in whatever APOE genetic population examined [48], not confirming the preliminary observation

made in the phase 2 studies. The overall criticism that might be formulated (retrospectively) here is that two elements that may increase the chance of success in a clinical study, a precise biological hypothesis and a quantitative assessable biomarker, were lacking.

 $\gamma$ -secretase is a protease complex that cleaves proteins at residues within their single membrane spanning domain. The most known substrate of  $\gamma$ -secretase is APP, whose cleavage produces A $\beta$ . The  $\gamma$ -secretase complex consists of four individual proteins, presenilin, nicastrin, APH-1 and PEN-2 [49]. A fifth protein, known as CD147, acts as a negative regulator of the complex [50]. Presenilin is the catalytic subunit and mutations in the presenilin gene rep-

resent a major genetic risk factor for AD [51]. Although γ-secretase mutations that completely knock out enzyme function prevent generation of Aβ, mutations that only partially knock out enzyme function often enhance generation of A $\beta$ , a finding ascribed to a gain of function [52]. Hence,  $\gamma$ -secretase inhibitors may enhance the production of A $\beta_{42}$  while blocking other  $\gamma$ -secretase activities, thus mimicking the effects of PS mutations [52], which may, at least in theory, produce paradoxical outcomes in AD trials (increased AB deposition and cognitive worsening). Furthermore, development of  $\gamma$ -secretase inhibitors as disease modifying drugs presents problems related to potential non-specific effects. This is because  $\gamma$ -secretase is not only responsible for AB generation but is also involved in intramembranous cleavage of several proteins, including the Notch receptor, ErbB4, p75NTR neurotrophin receptor, N-cadherin and the sodium channel β4 subunit [53]. Semagacestat was the first  $\gamma$ -secretase inhibitor to undergo extensive clinical testing and was shown to reduce Aβ concentrations in plasma and AB production in the central nervous system (CNS) [54,55]. Two large phase 3 RCTs with semagacestat were prematurely stopped because of some serious collateral adverse effects, including haematological, gastrointestinal and skin toxicity, that have been attributed to inhibition of the Notch signaling pathway [56]. Furthermore, in these studies, no improvement or moderate worsening of cognition was observed, perhaps related to  $\gamma$ -secretase inhibition within the CNS [57–59]. Notchsparing γ-secretase inhibitors (second generation inhibitors) and/or modulators (agents that shift γ-secretase cleavage activity from longer to shorter  $\beta$ -amyloid species, without affecting Notch cleavage) are in clinical development. Begacestat [60], BMS-708163 [61], PF-3084014 [62] and CHF-5074 [63] display a 10-100 fold selectivity on APP over the Notch cleavage. Some non-steroidal antiinflammatory drugs (NSAIDs) act as γ-secretase modulators, decreasing  $A\beta_{40}$  and  $A\beta_{42}$ , while increasing  $A\beta_{38}$  [53]. Tarenflurbil (the R-enantiomer of flurbiprofen) was tested in phase 3 RCTs but did not appear to slow cognitive decline [19], while increasing frequency of dizziness, anaemia, and infection [19]. The failure of tarenflurbil may be ascribed to low potency and poor brain penetration [64]. Furthermore, cyclo-oxygenase inhibition in microglia may result in inhibition of A $\beta$  clearance [65].

Other drugs, such as 1,4-dihydropyridine (DHP) L-type calcium channel blockers, are known to interfere with A $\beta$  production. Different large population-based studies have demonstrated that certain DHP calcium channel blockers used for the treatment of hypertension, such as nilvadipine, can reduce the risk of developing AD [66, 67]. Recent studies suggest that such benefits are not related to the drug's blood pressure lowering function [68]. Both nilvadipine and amlodipine decrease A $\beta$  production from APP in vitro, but only chronic oral treatment with nilvadipine reduces A $\beta$  accumulation in a transgenic model of AD, by targeting both production and clearance of A $\beta$  across the

BBB [68]. In a small study, nilvadipine slowed cognitive decline in MCI patients with hypertension [69]. Nilvadipine stabilizes cognition [70] and is well tolerated, with no dangerous blood pressure lowering effects [70]. A multicentre phase 3 clinical trial will start in January 2012 to assess the efficacy of nilvadipine as a disease modifying drug in AD patients (http://www.alzforum.org/new/detail.asp?id=2838).

#### Drugs to prevent Aβ aggregation

Aggregation of monomeric A $\beta$  species into higher molecular weight oligomers produces the primary neurotoxic species in AD [71, 72]. Tramiprosate (3-amino-L-propanesulfonic acid) is a glycosaminoglycan that binds to A $\beta$  monomers and prevents formation of oligomers, thus enhancing A $\beta$  clearance from the brain [73]. An initial, phase 2 study showed that tramiprosate reduces A $\beta_{42}$  concentrations in CSF [74]. In a larger, phase 3 study, however, tramiprosate did not determine clinical improvement [75], although a recent subanalysis suggests that it may exert some beneficial effects on memory, language and praxis skills [76], requiring further clinical evaluation.

Because zinc and copper are catalysts for A $\beta$  aggregation and stabilization of amyloid plaques, chelating agents may be effective in dissolving amyloid deposits *in vitro* and *in vivo*. PBT2 is an 8-hydroxy quinolone, orally administered and with good BBB permeability, that removes copper and zinc from CSF, promotes A $\beta$  oligomer clearance and restores cognition in AD mouse models [77, 78]. In a recent phase 2a study, PBT2 lowered A $\beta$ <sub>42</sub> in CSF and improved cognition, but no correlation was found between A $\beta$  in CSF and cognitive changes [77].

Scyllo-inositol (scyllo-cyclohexanehexol, AZD-103, ELND-005) is an orally administered stereoisomer of inositol that crosses the BBB using inositol transporters. Scyllo-inositol can directly bind to Aβ oligomers promoting dissociation of Aβ aggregates [79, 80]. Interestingly, TgCRND8 mice treated with AZD-103 show a 25% reduction of Aβ oligomers with a concomitant increase in monomeric species (+133%), suggesting that this drug can prevent the transition from A $\beta$  monomers to A $\beta$  oligomers [80]. Recently, a phase 2 clinical trial (NCT00568776) evaluating safety, efficacy and effects on biomarkers of ELND-005 in mild to moderate AD patients has been completed [81]. Of the three tested doses, 250, 1000 and 2000 mg, only 250 was well tolerated, whereas side effects in the two higher dose groups led to early discontinuation. In spite of lack of significant clinical improvement, patients receiving 250 mg of ELND005 had an increase in their brain ventricular volume as well as a reduction in CSF  $A\beta_{42}$ . Large-scale phase 3 clinical studies are needed to evaluate the clinical efficacy of ELND005.

Additional small molecules, including polyphenolic compounds such as curcumin (–)-epigallocatechin-3-



gallate (EGCG) and grape seed extract, attenuate  $A\beta$  aggregation [80, 82]. EGCG has shown good tolerability (NCT00525668) and is currently being evaluated in a phase 2–3 RCT (NCT00951834).

#### **Drugs to promote A**β clearance

Immunotherapy toward A $\beta$  is considered one of the most promising approaches to develop disease modifying drugs in AD, because it can potentially affect production, aggregation and deposition of AB [83, 84]. Active immunization by vaccination promotes formation of antibodies against pathogenic forms of AB, by stimulating an immune response, whereas passive immunotherapy supplies antibodies from an exogenous source [83]. Active Aß immunotherapy has been studied and validated since 1999, when it was demonstrated that generation of  $A\beta$  antibodies resulted in clearance of cerebral Aβ by microglial phagocytosis of antibody-opsonized Aβ deposits [85]. Aβ immunotherapy improves cognitive deficits in AD models and lowers plague load in non-human primates. Unfortunately, a phase 2 clinical trial of active immunization using full length human  $A\beta_{42}$  peptide with QS-21 adjuvant was stopped prematurely because some patients developed brain inflammation with aseptic meningoencephalitis [86]. T cell recognition of the human full length Aβ peptide may have induced an adverse autoimmune response [87]. Furthermore, although Aβ-specific antibodies clear brain amyloid plagues, they do not halt progressive neurodegeneration [88, 89] or affect vascular amyloid and hyperphosphorylated tau deposits [90]. Recent alternative approaches are based on shorter  $A\beta$  immunogens that target the N-terminus (strong B cell epitope) without affecting the mid-region and C-terminus (T cell epitopes) [84]. Because of the low responsiveness and adverse reactions to vaccines, passive immunotherapy has been proposed as an alternative strategy [91-93]. Passive immunotherapy, however, may also be associated with adverse effects such as vasogenic oedema and cerebral amyloid angiopathy with microhaemorrhages [5, 94, 95]. The most studied and advanced AB targeted antibody is bapineuzumab [96]. The efficacy and safety of bapineuzumab seem to be related to APOE allele status. In APOE ε4 carriers this drug can favour the onset of vasogenic oedema [93] that may limit its clinical use and has led to the abandonment of the highest dose of the drug (2 mg kg<sup>-1</sup>) [97]. Lower doses of bapineuzumab are currently used in phase 3 trials in ε4 carriers, whereas slightly higher doses can be used in non- £4 carriers [97]. To date seven phase 3 studies with bapineuzumab are ongoing (NCT00996918, NCT00574132, NCT00676143, NCT00667816, NCT00575055, NCT00998764 NCT00937352). Another humanized anti-Aβ monoclonal antibody in advanced clinical development is solanezumab; three phase 3 trials are ongoing (NCT 01127633,

NCT 00904683 and NCT00905372). Others antibodies in phase 1 and 2 trials include PF-04360365, GSK-933776, R-1415 and MABT-5102A.

Intravenous immunoglobulins (IVIG) contain naturally occurring autoantibodies that specifically recognize AB and block is toxic effects [98-101]. A phase 3 study with IVIG 10% is ongoing. Adekar et al. [102] showed that free human Igy heavy chains (HC) possess anti-amyloidogenic activity because they bind to an amyloid, fibril-related, conformational epitope while not affecting native Aβ monomers. Free human Igy HC offer the advantage of crossing the BBB and being less prone to adverse inflammatory side effects [103]. New strategies in the immunotherapy of AD should be directed to AB dimers and/or other toxic oligomers, preserving  $A\beta$  monomers, that may be involved in maintaining learning memory and neuronal survival [104]. Conformation specific antibodies, binding toxic AB oligomers without affecting A $\beta$  monomers, have been recently developed [105].

#### Strategies targeting tau

NFTs are intracellular aggregates of paired helical filaments whose main constituent is a hyperphosphorylated form of the protein tau [106]. Expression pattern of NFTs correlates with the clinical onset and progression of AD [107]. Although  $A\beta$  and tau have been considered for years as distinct with regard to their role in AD pathogenesis, recent evidence suggests that these two proteins significantly interact and that tau-related events are essential for AD pathogenesis [108]. Findings obtained in a triple transgenic mouse model of AD [109, 110], suggest that the two major histopathological hallmarks of AD, i.e. Aβ deposits and NFT, containing hyperphosphorylated tau, lie along the same pathological cascade. Aβ accumulation precedes and drives tau hyperphosphorylation via the activation of different kinases such as cyclin dependent kinase 5 (CDK5) and glycogen synthase kinase 3β (GSK3β) [108, 109, 111, 112]. Tau hyperphosphorylation leads to destabilization of neuronal microtubular dynamics, which finally results in an impairment of synaptic function [106]. The critical role of tau in mediating Aβ-induced neurodegeneration has been demonstrated both in in vitro and in vivo models [113, 114]. Tau hyperphosphorylation and subsequent accumulation in the dendritic compartment increases the vulnerability of neurons to the toxic effects of AB [108, 115]. Recent efforts in drug discovery have been therefore directed to develop inhibitors of tau-phosphorylation and compounds that prevent tau aggregation and/or promote disassembly. GSK3\beta is the main enzyme involved in tau hyperphosphorylation [116]. Lithium and valproate, currently used as mood stabilizers, both inhibit GSK3β and reduce tau phosphorylation in animal models [117]. Valproate has been studied in the Alzheimer's Disease Cooperative Study (ADCS) [118]. In this study valproate did not

modify cognition and functional status but reduced agitation and psychosis [118]. A more recent meta-analysis, however, shows that valproate is ineffective against agitation in demented patients, and is also associated with an unacceptable rate of adverse effects, such as falls, infection and gastrointestinal disorders [119]. Lithium is neuroprotective in animal models of AD, not only via the inhibition of GSK-3β, but also through other mechanisms, including reduction of Aβ production [120, 121] and release of TGF-β1 [122]. In patients treated with lithium for psychiatric disorders, the risk of developing AD is reduced [123, 124]. Some studies in AD patients, however, have failed to demonstrate a positive effect of lithium on cognitive performance [5, 125, 126]. A recent single centre study showed that lithium reduced both cognitive decline and CSF concentration of P-tau in patients with amnestic MCI [127]. Safety problems related to lithium treatment in elderly people need specific attention and may lead to high discontinuation rates in AD patients [128]. Other inhibitors of GSK-3β have shown neuroprotective effects in preclinical models of AD [129]. A phase 2 RCT has been recently completed with NP031112 (NCT00948259).

Methylthioninium chloride (MTC), also known as methylene blue, is a promising compound which possesses antioxidative properties, reduces A $\beta$  oligomerization and, most importantly, binds to the domain responsible for tau aggregation [130]. A phase 2b RCT study of MTC monotherapy in patients with mild to moderate AD showed improvement of cognition [131], that awaits to be validated in a forthcoming large scale phase 3 clinical trial [131].

### Other potential therapeutic approaches

A causal link between an impairment of nerve growth factor (NGF) pathway, activation of the amyloidogenic pathway and neurodegeneration in the AD brain has been proposed [132]. Targeted delivery of NGF to basal forebrain cholinergic neurons improves cognitive function in animal models of AD [132]. However, because protein growth factors do not cross the BBB, strategies targeting neurotrophic factors have been poorly exploited so far. Early studies, based on intracerebroventricular (ICV) infusion of NGF, showed a positive effect on cognitive function but were hampered by severe adverse effects related to ICV administration [133, 134]. To overcome these problems, the implant of autologous fibroblasts, genetically modified to express NGF into selected areas of CNS, has recently been proposed [135]. Other strategies use NGF gene-delivery through viral vectors [136-138] (NCT00876863 and NCT00087789). Encapsulated cell bio-delivery (ECB) provides NGF to cholinergic basal forebrain neurons through the stereotactic implantation of a catheter-like device containing NGF-producing cells (NsG0202). Preliminary results suggest a good safety and tolerability of NsG0202 [139].

Aβ triggers mitochondrial dysfunction through a number of pathways [140]. Rescue of mitochondrial function has been therefore considered as a new target to develop disease modifying drugs [141]. Latrepirdine is a weak inhibitor of cholinesterases and a low-affinity NMDA receptor antagonist, which exerts its neuroprotective effects through the stabilization of mitochondria via inhibition of mitochondrial permeability transition pores induced by A $\beta$  [142]. However, the ability of latrepirdine to improve cognition in AD is controversial, due to a discrepancy between the positive signal reported in a phase 2 clinical trial [143] and the subsequent null effect observed in a phase 3 trial [144]. Two RCTs are ongoing to assess the clinical efficacy of latrepirdine in combination with donepezil and memantine (NCT00829374 and NCT00912288). EGCG, mentioned above as an inhibitor of A $\beta$  aggregation, may also inhibit the release of apoptosis-inducing factor (AIF) from mitochondria [145].

Finally a new pharmacological target proposed for developing neuroprotective drugs in AD is the receptor for advanced glycation endproducts (RAGE), a transmembrane protein that belongs to the immunoglobulin superfamily localized in neurons, microglia, astrocytes and the BBB [146]. RAGE mediates the effects of Aβ on microglia, the BBB and neurons through different signaling pathways. RAGE enhances generation and accumulation of Aβ in the CNS by modulating BACE1 [147] and also promotes the transport of  $A\beta$  from vascular circulation to the brain. Data from autopsy brain tissues, in vitro cell cultures and transgenic mouse models suggest that the Aβ-RAGE interaction exaggerates neuronal stress, impairs learning memory and induces neuroinflammation [148]. A phase 2 trial with PF04494700, a RAGE antagonist, has been recently completed in mild to moderate AD patients and results on clinical efficacy of this drug are awaited in the next months.

Deep brain stimulation (DBS) of memory circuits has been proposed as an alternative, non-pharmacological approach for AD treatment [149]. A recent phase I trial conducted in six mild AD patients, receiving continuous stimulation for 12 months, suggests that DBS can revert impaired glucose utilization in the temporal and parietal lobes as assessed by PET and also slows cognitive decline. Additional studies are needed to confirm these preliminary results.

#### **Limitations and future directions**

The pharmacological treatment of AD actually involves cholinesterase inhibitors and memantine, which provide mainly symptomatic short term benefits without counteracting the progression of the disease. Drug discovery in AD has attempted in the last decade to develop disease



modifying drugs with the help of preclinical models, but none of these drugs has succeeded in phase 3. Factors that might explain this failure include suboptimal study design (lack and/or inadequate biomarkers and outcome measurements) and, most importantly, time course of treatment in relation to the development of disease. Available data from failed phase 3 studies suggest that mild to moderate AD patients may be too late in the disease process to improve substantively their outcome following drug treatment.

New criteria for the diagnosis of AD have enlarged the window for the detection of the early stages of the disease and include biomarkers mechanistically related to AD pathology. Adoption of these early biomarkers in implementing design of future studies is highly desirable. Finally, the heterogeneity of AD should be considered in the future when planning RCTs to evaluate the efficacy of disease modifying drugs. Because AD is heterogeneous in terms of clinical presentation, diagnostic issues, underlying neuropathology and mixed causes of dementia have been described in many late onset AD patients, a major challenge will be to identify subgroups with homogeneous biomarkers and to improve the neuropsychological tools for detecting deficits of episodic memory in amnestic MCI patients at high risk to convert into AD. At present, the focus in AD drug development is shifting from treatment to prevention [150]. The new strategy will examine the potential neuroprotective activity of disease modifying drugs in the presymptomatic stages of AD, with the help of biomarkers that predict disease progression before development of overt dementia.

#### **Competing Interests**

There are no competing interests to declare.

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#### **REFERENCES**

- 1 Ballard C, Day S, Sharp S, Wing G, Sorensen S. Neuropsychiatric symptoms in dementia: importance and treatment considerations. Int Rev Psychiatry 2008; 20: 396–404.
- **2** Hardy J. The amyloid hypothesis for Alzheimer's disease: a critical reappraisal. J Neurochem 2009; 110: 1129–34.
- **3** Cerpa W, Dinamarca MC, Inestrosa NC. Structure-function implications in Alzheimer's disease: effect of Abeta oligomers at central synapses. Curr Alzheimer Res 2008; 5: 233–43.
- **4** Klafki HW, Staufenbiel M, Kornhuber J, Wiltfang J. Therapeutic approaches to Alzheimer's disease. Brain 2006; 129: 2840–55.

- **5** Mangialasche F, Solomon A, Winblad B, Mecocci P, Kivipelto M. Alzheimer's disease: clinical trials and drug development. Lancet Neurol 2010; 9: 702–16.
- **6** Nordberg A. Mechanisms behind the neuroprotective actions of cholinesterase inhibitors in Alzheimer disease. Alzheimer Dis Assoc Disord 2006; 20: \$12–8.
- 7 Wu HM, Tzeng NS, Qian L, Wei SJ, Hu X, Chen SH, Rawls SM, Flood P, Hong JS, Lu RB. Novel neuroprotective mechanisms of memantine: increase in neurotrophic factor release from astroglia and anti-inflammation by preventing microglial activation. Neuropsychopharmacology 2009; 34: 2344–57.
- 8 McKhann GM, Knopman DS, Chertkow H, Hyman BT, Jack CR Jr, Kawas CH, Klunk WE, Koroshetz WJ, Manly JJ, Mayeux R, Mohs RC, Morris JC, Rossor MN, Scheltens P, Carrillo MC, Thies B, Weintraub S, Phelps CH. The diagnosis of dementia due to Alzheimer's disease: recommendations from the National Institute on Aging-Alzheimer's Association workgroups on diagnostic guidelines for Alzheimer's disease. Alzheimers Dement 2011; 7: 263–9.
- 9 Albert MS, DeKosky ST, Dickson D, Dubois B, Feldman HH, Fox NC, Gamst A, Holtzman DM, Jagust WJ, Petersen RC, Snyder PJ, Carrillo MC, Thies B, Phelps CH. The diagnosis of mild cognitive impairment due to Alzheimer's disease: recommendations from the National Institute on Aging-Alzheimer's Association workgroups on diagnostic guidelines for Alzheimer's disease. Alzheimers Dement 2011; 7: 270–9.
- 10 Sperling RA, Aisen PS, Beckett LA, Bennett DA, Craft S, Fagan AM, Iwatsubo T, Jack CR Jr, Kaye J, Montine TJ, Park DC, Reiman EM, Rowe CC, Siemers E, Stern Y, Yaffe K, Carrillo MC, Thies B, Morrison-Bogorad M, Wagster MV, Phelps CH. Toward defining the preclinical stages of Alzheimer's disease: recommendations from the National Institute on Aging-Alzheimer's Association workgroups on diagnostic guidelines for Alzheimer's disease. Alzheimers Dement 2011; 7: 280–92.
- **11** Siemers ER. How can we recognize 'disease modification' effects? J Nutr Health Aging 2009; 13: 341–3.
- **12** Cummings JL. Biomarkers in Alzheimer's disease drug development. Alzheimers Dement 2011; 7: e13–44.
- 13 Fagan AM, Mintun MA, Mach RH, Lee SY, Dence CS, Shah AR, LaRossa GN, Spinner ML, Klunk WE, Mathis CA, DeKosky ST, Morris JC, Holtzman DM. Inverse relation between *in vivo* amyloid imaging load and cerebrospinal fluid Abeta42 in humans. Ann Neurol 2006; 59: 512–9.
- **14** Ringman JM, Taylor K, Teng E, Coppola G, Gylys K. Longitudinal change in CSF biomarkers in a presymptomatic carrier of an APP mutation. Neurology 2011; 76: 2124–5.
- 15 van der Worp HB, Macleod MR. Preclinical studies of human disease: time to take methodological quality seriously. J Mol Cell Cardiol 2011; 51: 449–50.
- **16** Dirnagl U. Bench to bedside: the quest for quality in experimental stroke research. J Cereb Blood Flow Metab 2006; 26: 1465–78.

- 17 Alzheimer A, Stelzmann RA, Schnitzlein HN, Murtagh FR. An English translation of Alzheimer's 1907 paper, 'Uber eine eigenartige Erkankung der Hirnrinde'. Clin Anat 1995; 8: 429–31.
- **18** Extance A. Alzheimer's failure raises questions about disease-modifying strategies. Nat Rev Drug Discov 2010; 9: 749–51.
- 19 Green RC, Schneider LS, Amato DA, Beelen AP, Wilcock G, Swabb EA, Zavitz KH. Effect of tarenflurbil on cognitive decline and activities of daily living in patients with mild Alzheimer disease: a randomized controlled trial. JAMA 2009: 302: 2557–64.
- 20 Puzzo D, Privitera L, Fa M, Staniszewski A, Hashimoto G, Aziz F, Sakurai M, Ribe EM, Troy CM, Mercken M, Jung SS, Palmeri A, Arancio O. Endogenous amyloid-beta is necessary for hippocampal synaptic plasticity and memory. Ann Neurol 2011; 69: 819–30.
- 21 Malchiodi-Albedi F, Paradisi S, Matteucci A, Frank C, Diociaiuti M. Amyloid oligomer neurotoxicity, calcium dysregulation, and lipid rafts. Int. J Alzheimers Dis 2011; DOI: 10.4061/2011/906964.
- **22** Galvin JE. Dementia screening, biomarkers and protein misfolding: implications for public health and diagnosis. Prion 2011; 5: 16–21.
- 23 Grasso G. The use of mass spectrometry to study amyloid-beta peptides. Mass Spectrom Rev 2011; 30: 347–65.
- 24 Lichtenthaler SF, Haass C, Steiner H. Regulated intramembrane proteolysis lessons from amyloid precursor protein processing. J Neurochem 2011; 117: 779–96.
- **25** Eder J, Hommel U, Cumin F, Martoglio B, Gerhartz B. Aspartic proteases in drug discovery. Curr Pharm Des 2007; 13: 271–85.
- 26 Zetterberg H, Andreasson U, Hansson O, Wu G, Sankaranarayanan S, Andersson ME, Buchhave P, Londos E, Umek RM, Minthon L, Simon AJ, Blennow K. Elevated cerebrospinal fluid BACE1 activity in incipient Alzheimer disease. Arch Neurol 2008; 65: 1102–7.
- 27 Wang S, Jia J. Promoter polymorphisms which modulate BACE1 expression are associated with sporadic Alzheimer's disease. Am J Med Genet B Neuropsychiatr Genet 2010; 153B: 159–66.
- 28 Ohno M, Sametsky EA, Younkin LH, Oakley H, Younkin SG, Citron M, Vassar R, Disterhoft JF. BACE1 deficiency rescues memory deficits and cholinergic dysfunction in a mouse model of Alzheimer's disease. Neuron 2004; 41: 27–33.
- 29 Landreth G, Jiang Q, Mandrekar S, Heneka M. PPARgamma agonists as therapeutics for the treatment of Alzheimer's disease. Neurother 2008; 5: 481–9.
- **30** Chang WP, Huang X, Downs D, Cirrito JR, Koelsch G, Holtzman DM, Ghosh AK, Tang J. Beta-secretase inhibitor GRL-8234 rescues age-related cognitive decline in APP transgenic mice. FASEB J 2011; 25: 775–84.
- **31** Ghosh AK, Kumaragurubaran N, Hong L, Koelsh G, Tang J. Memapsin 2 (beta-secretase) inhibitors: drug development. Curr Alzheimer Res 2008; 5: 121–31.

- **32** Albert JS. Progress in the development of beta-secretase inhibitors for Alzheimer's disease. Prog Med Chem 2009; 48: 133–61.
- **33** Francis GA, Fayard E, Picard F, Auwerx J. Nuclear receptors and the control of metabolism. Annu Rev Physiol 2003; 65: 261–311.
- **34** Akter K, Lanza EA, Martin SA, Myronyuk N, Rua M, Raffa RB. Diabetes mellitus and Alzheimer's disease: shared pathology and treatment? Br J Clin Pharmacol 2011; 71: 365–76.
- **35** Peila R, Rodriguez BL, Launer LJ. Type 2 diabetes, APOE gene, and the risk for dementia and related pathologies: the Honolulu-Asia Aging Study. Diabetes 2002; 51: 1256–62.
- **36** Pedersen WA, McMillan PJ, Kulstad JJ, Leverenz JB, Craft S, Haynatzki GR. Rosiglitazone attenuates learning and memory deficits in Tg2576 Alzheimer mice. Exp Neurol 2006: 199: 265–73.
- **37** Selkoe DJ. Clearing the brain's amyloid cobwebs. Neuron 2001; 32: 177–80.
- **38** Du J, Zhang L, Liu S, Zhang C, Huang X, Li J, Zhao N, Wang Z. PPARgamma transcriptionally regulates the expression of insulin-degrading enzyme in primary neurons. Biochem Biophys Res Commun 2009; 383: 485–90.
- **39** Kalinin S, Richardson JC, Feinstein DL. A PPARdelta agonist reduces amyloid burden and brain inflammation in a transgenic mouse model of Alzheimer's disease. Curr Alzheimer Res 2009: 6: 431–7.
- 40 Watson GS, Cholerton BA, Reger MA, Baker LD, Plymate SR, Asthana S, Fishel MA, Kulstad JJ, Green PS, Cook DG, Kahn SE, Keeling ML, Craft S. Preserved cognition in patients with early Alzheimer disease and amnestic mild cognitive impairment during treatment with rosiglitazone: a preliminary study. Am J Geriatr Psychiatry 2005; 13: 950–8.
- **41** Strozyk D, Blennow K, White LR, Launer LJ. CSF Abeta 42 levels correlate with amyloid-neuropathology in a population-based autopsy study. Neurology 2003; 60: 652–6.
- **42** Irizarry MC. Biomarkers of Alzheimer disease in plasma. NeuroRx 2004; 1: 226–34.
- **43** Vanderstichele H, Van Kerschaver E, Hesse C, Davidsson P, Buyse MA, Andreasen N, Minthon L, Wallin A, Blennow K, Vanmechelen E. Standardization of measurement of beta-amyloid(1-42) in cerebrospinal fluid and plasma. Amyloid 2000; 7: 245–58.
- **44** Risner ME, Saunders AM, Altman JF, Ormandy GC, Craft S, Foley IM, Zvartau-Hind ME, Hosford DA, Roses AD. Efficacy of rosiglitazone in a genetically defined population with mild-to-moderate Alzheimer's disease. Pharmacogenomics J 2006; 6: 246–54.
- **45** Reiman EM, Caselli RJ, Yun LS, Chen K, Bandy D, Minoshima S, Thibodeau SN, Osborne D. Preclinical evidence of Alzheimer's disease in persons homozygous for the epsilon 4 allele for apolipoprotein E. N Engl J Med 1996; 334: 752–8.



- **46** Gibson GE, Haroutunian V, Zhang H, Park LC, Shi Q, Lesser M, Mohs RC, Sheu RK, Blass JP. Mitochondrial damage in Alzheimer's disease varies with apolipoprotein E genotype. Ann Neurol 2000; 48: 297–303.
- 47 Bubber P, Haroutunian V, Fisch G, Blass JP, Gibson GE. Mitochondrial abnormalities in Alzheimer brain: mechanistic implications. Ann Neurol 2005; 57: 695–703.
- **48** Gold M, Alderton C, Zvartau-Hind M, Egginton S, Saunders AM, Irizarry M, Craft S, Landreth G, Linnamagi U, Sawchak S. Rosiglitazone monotherapy in mild-to-moderate Alzheimer's disease: results from a randomized, double-blind, placebo-controlled phase III study. Dement Geriatr Cogn Disord 2010; 30: 131–46.
- **49** Kaether C, Haass C, Steiner H. Assembly, trafficking and function of gamma-secretase. Neurodegener Dis 2006; 3: 275–83.
- 50 Zhou S, Zhou H, Walian PJ, Jap BK. The discovery and role of CD147 as a subunit of gamma-secretase complex. Drug News Perspect 2006; 19: 133–8.
- 51 Chen F, Hasegawa H, Schmitt-Ulms G, Kawarai T, Bohm C, Katayama T, Gu Y, Sanjo N, Glista M, Rogaeva E, Wakutani Y, Pardossi-Piquard R, Ruan X, Tandon A, Checler F, Marambaud P, Hansen K, Westaway D, St George-Hyslop P, Fraser P. TMP21 is a presenilin complex component that modulates gamma-secretase but not epsilon-secretase activity. Nature 2006; 440: 1208–12.
- **52** Shen J, Kelleher RJ 3rd. The presentilin hypothesis of Alzheimer's disease: evidence for a loss-of-function pathogenic mechanism. Proc Natl Acad Sci U S A 2007; 104: 403–9.
- 53 Tomita T. Secretase inhibitors and modulators for Alzheimer's disease treatment. Expert Rev Neurother 2009; 9: 661–79
- **54** Henley DB, May PC, Dean RA, Siemers ER. Development of semagacestat (LY450139), a functional gamma-secretase inhibitor, for the treatment of Alzheimer's disease. Expert Opin Pharmacother 2009; 10: 1657–64.
- 55 Bateman RJ, Siemers ER, Mawuenyega KG, Wen G, Browning KR, Sigurdson WC, Yarasheski KE, Friedrich SW, Demattos RB, May PC, Paul SM, Holtzman DM. A gamma-secretase inhibitor decreases amyloid-beta production in the central nervous system. Ann Neurol 2009; 66: 48–54.
- **56** Schor NF. What the halted phase III gamma-secretase inhibitor trial may (or may not) be telling us. Ann Neurol 2011; 69: 237–9.
- **57** Yan C, Liang Y, Nylander KD, Wong J, Rudavsky RM, Saragovi HU, Schor NF. p75-nerve growth factor as an antiapoptotic complex: independence *versus* cooperativity in protection from enediyne chemotherapeutic agents. Mol Pharmacol 2002: 61: 710–9.
- 58 Yan C, Mirnics ZK, Portugal CF, Liang Y, Nylander KD, Rudzinski M, Zaccaro C, Saragovi HU, Schor NF. Cholesterol biosynthesis and the pro-apoptotic effects of the p75 nerve growth factor receptor in PC12 pheochromocytoma cells. Brain Res Mol Brain Res 2005; 139: 225–34.

- 59 Rabizadeh S, Bitler CM, Butcher LL, Bredesen DE. Expression of the low-affinity nerve growth factor receptor enhances beta-amyloid peptide toxicity. Proc Natl Acad Sci U S A 1994; 91: 10703–6.
- 60 Martone RL, Zhou H, Atchison K, Comery T, Xu JZ, Huang X, Gong X, Jin M, Kreft A, Harrison B, Mayer SC, Aschmies S, Gonzales C, Zaleska MM, Riddell DR, Wagner E, Lu P, Sun SC, Sonnenberg-Reines J, Oganesian A, Adkins K, Leach MW, Clarke DW, Huryn D, Abou-Gharbia M, Magolda R, Bard J, Frick G, Raje S, Forlow SB, Balliet C, Burczynski ME, Reinhart PH, Wan HI, Pangalos MN, Jacobsen JS. Begacestat (GSI-953): a novel, selective thiophene sulfonamide inhibitor of amyloid precursor protein gamma-secretase for the treatment of Alzheimer's disease. J Pharmacol Exp Ther 2009; 331: 598–608.
- **61** Imbimbo BP. Therapeutic potential of gamma-secretase inhibitors and modulators. Curr Top Med Chem 2008; 8: 54–61.
- **62** Lanz TA, Wood KM, Richter KE, Nolan CE, Becker SL, Pozdnyakov N, Martin BA, Du P, Oborski CE, Wood DE, Brown TM, Finley JE, Sokolowski SA, Hicks CD, Coffman KJ, Geoghegan KF, Brodney MA, Liston D, Tate B. Pharmacodynamics and pharmacokinetics of the gamma-secretase inhibitor PF-3084014. J Pharmacol Exp Ther 2010; 334: 269–77.
- 63 Imbimbo BP, Hutter-Paier B, Villetti G, Facchinetti F, Cenacchi V, Volta R, Lanzillotta A, Pizzi M, Windisch M. CHF5074, a novel gamma-secretase modulator, attenuates brain beta-amyloid pathology and learning deficit in a mouse model of Alzheimer's disease. Br J Pharmacol 2009; 156: 982–93.
- **64** Imbimbo BP, Giardina GA. Gamma-secretase inhibitors and modulators for the treatment of Alzheimer's disease: disappointments and hopes. Curr Top Med Chem 2011; 11: 1555–70.
- **65** Persaud-Sawin DA, Banach L, Harry GJ. Raft aggregation with specific receptor recruitment is required for microglial phagocytosis of Abeta42. Glia 2009; 57: 320–35.
- 66 Forette F, Seux ML, Staessen JA, Thijs L, Babarskiene MR, Babeanu S, Bossini A, Fagard R, Gil-Extremera B, Laks T, Kobalava Z, Sarti C, Tuomilehto J, Vanhanen H, Webster J, Yodfat Y, Birkenhager WH. The prevention of dementia with antihypertensive treatment: new evidence from the Systolic Hypertension in Europe (Syst-Eur) study. Arch Intern Med 2002; 162: 2046–52.
- **67** Hanon O, Forette F. Prevention of dementia: lessons from SYST-EUR and PROGRESS. J Neurol Sci 2004; 226: 71–4.
- **68** Paris D, Bachmeier C, Patel N, Quadros A, Volmar CH, Laporte V, Ganey J, Beaulieu-Abdelahad D, Ait-Ghezala G, Crawford F, Mullan MJ. Selective antihypertensive dihydropyridines lower Abeta accumulation by targeting both the production and the clearance of Abeta across the blood-brain barrier. Mol Med 2011; 17: 149–62.
- **69** Hanyu H, Hirao K, Shimizu S, Sato T, Kiuchi A, Iwamoto T. Nilvadipine prevents cognitive decline of patients with mild cognitive impairment. Int J Geriatr Psychiatry 2007; 22: 1264–6.

- 70 Kennelly SP, Abdullah L, Paris D, Parish J, Mathura V, Mullan M, Crawford F, Lawlor BA, Kenny RA. Demonstration of safety in Alzheimer's patients for intervention with an anti-hypertensive drug nilvadipine: results from a 6-week open label study. Int J Geriatr Psychiatry 2011; 26: 1038–45.
- 71 Shankar GM, Bloodgood BL, Townsend M, Walsh DM, Selkoe DJ, Sabatini BL. Natural oligomers of the Alzheimer amyloid-beta protein induce reversible synapse loss by modulating an NMDA-type glutamate receptor-dependent signaling pathway. J Neurosci 2007; 27: 2866–75.
- 72 Walsh DM, Selkoe DJ. A beta oligomers a decade of discovery. J Neurochem 2007; 101: 1172–84.
- 73 Gervais F, Paquette J, Morissette C, Krzywkowski P, Yu M, Azzi M, Lacombe D, Kong X, Aman A, Laurin J, Szarek WA, Tremblay P. Targeting soluble Abeta peptide with Tramiprosate for the treatment of brain amyloidosis. Neurobiol Aging 2007; 28: 537–47.
- 74 Aisen PS, Saumier D, Briand R, Laurin J, Gervais F, Tremblay P, Garceau D. A Phase II study targeting amyloid-beta with 3APS in mild-to-moderate Alzheimer disease. Neurology 2006; 67: 1757–63.
- 75 Gauthier S, Aisen PS, Ferris SH, Saumier D, Duong A, Haine D, Garceau D, Suhy J, Oh J, Lau W, Sampalis J. Effect of tramiprosate in patients with mild-to-moderate Alzheimer's disease: exploratory analyses of the MRI sub-group of the Alphase study. J Nutr Health Aging 2009; 13: 550–7.
- **76** Saumier D, Duong A, Haine D, Garceau D, Sampalis J. Domain-specific cognitive effects of tramiprosate in patients with mild to moderate Alzheimer's disease: ADAS-cog subscale results from the Alphase Study. J Nutr Health Aging 2009; 13: 808–12.
- 77 Faux NG, Ritchie CW, Gunn A, Rembach A, Tsatsanis A, Bedo J, Harrison J, Lannfelt L, Blennow K, Zetterberg H, Ingelsson M, Masters CL, Tanzi RE, Cummings JL, Herd CM, Bush AI. PBT2 rapidly improves cognition in Alzheimer's disease: additional phase II analyses. J Alzheimers Dis 2010; 20: 509–16.
- 78 Adlard PA, Cherny RA, Finkelstein DI, Gautier E, Robb E, Cortes M, Volitakis I, Liu X, Smith JP, Perez K, Laughton K, Li QX, Charman SA, Nicolazzo JA, Wilkins S, Deleva K, Lynch T, Kok G, Ritchie CW, Tanzi RE, Cappai R, Masters CL, Barnham KJ, Bush AI. Rapid restoration of cognition in Alzheimer's transgenic mice with 8-hydroxy quinoline analogs is associated with decreased interstitial Abeta. Neuron 2008; 59: 43–55.
- 79 Fenili D, Brown M, Rappaport R, McLaurin J. Properties of scyllo-inositol as a therapeutic treatment of AD-like pathology. J Mol Med (Berl) 2007; 85: 603–11.
- 80 Amijee H, Scopes DI. The quest for small molecules as amyloid inhibiting therapies for Alzheimer's disease. J Alzheimers Dis 2009; 17: 33–47.
- **81** Salloway S, Sperling R, Keren R, Porsteinsson AP, van Dyck CH, Tariot PN, Gilman S, Arnold D, Abushakra S, Hernandez C, Crans G, Liang E, Quinn G, Bairu M, Pastrak A, Cedarbaum JM. A phase 2 randomized trial of ELND005, scyllo-inositol, in mild to moderate Alzheimer disease. Neurology 2011; 77: 1253–62.

- **82** Mandel SA, Amit T, Kalfon L, Reznichenko L, Weinreb O, Youdim MB. Cell signaling pathways and iron chelation in the neurorestorative activity of green tea polyphenols: special reference to epigallocatechin gallate (EGCG). J Alzheimers Dis 2008; 15: 211–22.
- 83 Town T. Alternative Abeta immunotherapy approaches for Alzheimer's disease. CNS Neurol Disord Drug Targets 2009; 8: 114–27.
- **84** Lemere CA. Developing novel immunogens for a safe and effective Alzheimer's disease vaccine. Prog Brain Res 2009; 175: 83–93.
- 85 Schenk D, Barbour R, Dunn W, Gordon G, Grajeda H, Guido T, Hu K, Huang J, Johnson-Wood K, Khan K, Kholodenko D, Lee M, Liao Z, Lieberburg I, Motter R, Mutter L, Soriano F, Shopp G, Vasquez N, Vandevert C, Walker S, Wogulis M, Yednock T, Games D, Seubert P. Immunization with amyloid-beta attenuates Alzheimer-disease-like pathology in the PDAPP mouse. Nature 1999; 400: 173–7.
- **86** Check E. Nerve inflammation halts trial for Alzheimer's drug. Nature 2002; 415: 462.
- **87** Wisniewski T, Frangione B. Immunological and anti-chaperone therapeutic approaches for Alzheimer disease. Brain Pathol 2005; 15: 72–7.
- **88** Nicoll JA, Wilkinson D, Holmes C, Steart P, Markham H, Weller RO. Neuropathology of human Alzheimer disease after immunization with amyloid-beta peptide: a case report. Nat Med 2003; 9: 448–52.
- 89 Holmes C, Boche D, Wilkinson D, Yadegarfar G, Hopkins V, Bayer A, Jones RW, Bullock R, Love S, Neal JW, Zotova E, Nicoll JA. Long-term effects of Abeta42 immunisation in Alzheimer's disease: follow-up of a randomised, placebo-controlled phase I trial. Lancet 2008; 372: 216–23.
- 90 Kokjohn TA, Roher AE. Antibody responses, amyloid-beta peptide remnants and clinical effects of AN-1792 immunization in patients with AD in an interrupted trial. CNS Neurol Disord Drug Targets 2009; 8: 88–97.
- 91 Bard F, Cannon C, Barbour R, Burke RL, Games D, Grajeda H, Guido T, Hu K, Huang J, Johnson-Wood K, Khan K, Kholodenko D, Lee M, Lieberburg I, Motter R, Nguyen M, Soriano F, Vasquez N, Weiss K, Welch B, Seubert P, Schenk D, Yednock T. Peripherally administered antibodies against amyloid beta-peptide enter the central nervous system and reduce pathology in a mouse model of Alzheimer disease. Nat Med 2000; 6: 916–9.
- **92** DeMattos RB, Bales KR, Cummins DJ, Dodart JC, Paul SM, Holtzman DM. Peripheral anti-A beta antibody alters CNS and plasma A beta clearance and decreases brain A beta burden in a mouse model of Alzheimer's disease. Proc Natl Acad Sci U S A 2001; 98: 8850–5.
- 93 Salloway S, Sperling R, Gilman S, Fox NC, Blennow K, Raskind M, Sabbagh M, Honig LS, Doody R, van Dyck CH, Mulnard R, Barakos J, Gregg KM, Liu E, Lieberburg I, Schenk D, Black R, Grundman M. A phase 2 multiple ascending dose trial of bapineuzumab in mild to moderate Alzheimer disease. Neurology 2009; 73: 2061–70.



- **94** Pfeifer LA, White LR, Ross GW, Petrovitch H, Launer LJ. Cerebral amyloid angiopathy and cognitive function: the HAAS autopsy study. Neurology 2002; 58: 1629–34.
- **95** Morgan D. Immunotherapy for Alzheimer's disease. J Intern Med 2011; 269: 54–63.
- **96** Kerchner GA, Boxer AL. Bapineuzumab. Expert Opin Biol Ther 2010; 10: 1121–30.
- 97 Panza F, Frisardi V, Imbimbo BP, Seripa D, Paris F, Santamato A, D'Onofrio G, Logroscino G, Pilotto A, Solfrizzi V. Anti-beta-amyloid immunotherapy for Alzheimer's disease: focus on bapineuzumab. Curr Alzheimer Res 2011; 8: 808–17.
- 98 Dodel R, Neff F, Noelker C, Pul R, Du Y, Bacher M, Oertel W. Intravenous immunoglobulins as a treatment for Alzheimer's disease: rationale and current evidence. Drugs 2010; 70: 513–28.
- 99 Magga J, Puli L, Pihlaja R, Kanninen K, Neulamaa S, Malm T, Hartig W, Grosche J, Goldsteins G, Tanila H, Koistinaho J, Koistinaho M. Human intravenous immunoglobulin provides protection against Abeta toxicity by multiple mechanisms in a mouse model of Alzheimer's disease. J Neuroinflammation 2010; DOI: 10.1186/1742-2094-7-90.
- 100 Dodel RC, Du Y, Depboylu C, Hampel H, Frolich L, Haag A, Hemmeter U, Paulsen S, Teipel SJ, Brettschneider S, Spottke A, Nolker C, Moller HJ, Wei X, Farlow M, Sommer N, Oertel WH. Intravenous immunoglobulins containing antibodies against beta-amyloid for the treatment of Alzheimer's disease. J Neurol Neurosurg Psychiatry 2004; 75: 1472–4.
- 101 Relkin NR, Szabo P, Adamiak B, Burgut T, Monthe C, Lent RW, Younkin S, Younkin L, Schiff R, Weksler ME. 18-month study of intravenous immunoglobulin for treatment of mild Alzheimer disease. Neurobiol Aging 2009; 30: 1728–36.
- **102** Adekar SP, Klyubin I, Macy S, Rowan MJ, Solomon A, Dessain SK, O'Nuallain B. Inherent anti-amyloidogenic activity of human immunoglobulin gamma heavy chains. J Biol Chem 2010; 285: 1066–74.
- 103 Robert R, Dolezal O, Waddington L, Hattarki MK, Cappai R, Masters CL, Hudson PJ, Wark KL. Engineered antibody intervention strategies for Alzheimer's disease and related dementias by targeting amyloid and toxic oligomers. Protein Eng Des Sel 2009; 22: 199–208.
- 104 Giuffrida ML, Caraci F, Pignataro B, Cataldo S, De Bona P, Bruno V, Molinaro G, Pappalardo G, Messina A, Palmigiano A, Garozzo D, Nicoletti F, Rizzarelli E, Copani A. Beta-amyloid monomers are neuroprotective. J Neurosci 2009; 29: 10582–7.
- **105** Lambert MP, Velasco PT, Viola KL, Klein WL. Targeting generation of antibodies specific to conformational epitopes of amyloid beta-derived neurotoxins. CNS Neurol Disord Drug Targets 2009; 8: 65–81.
- 106 Iqbal K, Alonso AC, Gong CX, Khatoon S, Pei JJ, Wang JZ, Grundke-Iqbal I. Mechanisms of neurofibrillary degeneration and the formation of neurofibrillary tangles. J Neural Transm Suppl 1998; 53: 169–80.

- 107 Bierer LM, Hof PR, Purohit DP, Carlin L, Schmeidler J, Davis KL, Perl DP. Neocortical neurofibrillary tangles correlate with dementia severity in Alzheimer's disease. Arch Neurol 1995; 52: 81–8.
- 108 Ittner LM, Gotz J. Amyloid-beta and tau a toxic pas de deux in Alzheimer's disease. Nat Rev Neurosci 2011; 12: 65–72.
- **109** Oddo S, Caccamo A, Shepherd JD, Murphy MP, Golde TE, Kayed R, Metherate R, Mattson MP, Akbari Y, LaFerla FM. Triple-transgenic model of Alzheimer's disease with plaques and tangles: intracellular Abeta and synaptic dysfunction. Neuron 2003; 39: 409–21.
- **110** Blurton-Jones M, Laferla FM. Pathways by which Abeta facilitates tau pathology. Curr Alzheimer Res 2006; 3: 437–48.
- **111** Gotz J, Chen F, van Dorpe J, Nitsch RM. Formation of neurofibrillary tangles in P301l tau transgenic mice induced by Abeta 42 fibrils. Science 2001; 293: 1491–5.
- 112 Caricasole A, Copani A, Caraci F, Aronica E, Rozemuller AJ, Caruso A, Storto M, Gaviraghi G, Terstappen GC, Nicoletti F. Induction of Dickkopf-1, a negative modulator of the Wnt pathway, is associated with neuronal degeneration in Alzheimer's brain. J Neurosci 2004; 24: 6021–7.
- **113** Rapoport M, Dawson HN, Binder LI, Vitek MP, Ferreira A. Tau is essential to beta-amyloid-induced neurotoxicity. Proc Natl Acad Sci U S A 2002: 99: 6364–9.
- **114** Terwel D, Muyllaert D, Dewachter I, Borghgraef P, Croes S, Devijver H, Van Leuven F. Amyloid activates GSK-3beta to aggravate neuronal tauopathy in bigenic mice. Am J Pathol 2008; 172: 786–98.
- 115 Gong CX, Iqbal K. Hyperphosphorylation of microtubule-associated protein tau: a promising therapeutic target for Alzheimer disease. Curr Med Chem 2008; 15: 2321–8.
- 116 Takashima A, Honda T, Yasutake K, Michel G, Murayama O, Murayama M, Ishiguro K, Yamaguchi H. Activation of tau protein kinase I/glycogen synthase kinase-3beta by amyloid beta peptide (25-35) enhances phosphorylation of tau in hippocampal neurons. Neurosci Res 1998; 31: 317–23.
- **117** Tariot PN, Aisen PS. Can lithium or valproate untie tangles in Alzheimer's disease? J Clin Psychiatry 2009; 70: 919–21.
- **118** Reiman EM, Langbaum JB, Tariot PN. Alzheimer's prevention initiative: a proposal to evaluate presymptomatic treatments as quickly as possible. Biomark Med 2010; 4: 3–14.
- **119** Lonergan E, Luxenberg J. Valproate preparations for agitation in dementia. Cochrane Database Syst Rev 2009; (3): CD003945.
- **120** Wada A, Yokoo H, Yanagita T, Kobayashi H. Lithium: potential therapeutics against acute brain injuries and chronic neurodegenerative diseases. J Pharmacol Sci 2005; 99: 307–21.
- **121** Lauterbach EC, Victoroff J, Coburn KL, Shillcutt SD, Doonan SM, Mendez MF. Psychopharmacological

- neuroprotection in neurodegenerative disease: assessing the preclinical data. J Neuropsychiatry Clin Neurosci 2010; 22: 8–18.
- **122** Caraci F, Battaglia G, Bruno V, Bosco P, Carbonaro V, Giuffrida ML, Drago F, Sortino MA, Nicoletti F, Copani A. TGF-beta1 pathway as a new target for neuroprotection in Alzheimer's disease. CNS Neurosci Ther 2011; 17: 237–49.
- **123** Nunes PV, Forlenza OV, Gattaz WF. Lithium and risk for Alzheimer's disease in elderly patients with bipolar disorder. Br J Psychiatry 2008; 190: 359–60.
- **124** Yeh HL, Tsai SJ. Lithium may be useful in the prevention of Alzheimer's disease in individuals at risk of presenile familial Alzheimer's disease. Med Hypotheses 2008; 71: 948–51.
- 125 Hampel H, Ewers M, Burger K, Annas P, Mortberg A, Bogstedt A, Frolich L, Schroder J, Schonknecht P, Riepe MW, Kraft I, Gasser T, Leyhe T, Moller HJ, Kurz A, Basun H. Lithium trial in Alzheimer's disease: a randomized, single-blind, placebo-controlled, multicenter 10-week study. J Clin Psychiatry 2009; 70: 922–31.
- **126** Vellas B, Andrieu S, Sampaio C, Wilcock G. Disease-modifying trials in Alzheimer's disease: a European task force consensus. Lancet Neurol 2007; 6: 56–62.
- 127 Forlenza OV, Diniz BS, Radanovic M, Santos FS, Talib LL, Gattaz WF. Disease-modifying properties of long-term lithium treatment for amnestic mild cognitive impairment: randomised controlled trial. Br J Psychiatry 2011; 198: 351–6.
- **128** Macdonald A, Briggs K, Poppe M, Higgins A, Velayudhan L, Lovestone S. A feasibility and tolerability study of lithium in Alzheimer's disease. Int J Geriatr Psychiatry 2008; 23: 704–11
- **129** Sereno L, Coma M, Rodriguez M, Sanchez-Ferrer P, Sanchez MB, Gich I, Agullo JM, Perez M, Avila J, Guardia-Laguarta C, Clarimon J, Lleo A, Gomez-Isla T. A novel GSK-3beta inhibitor reduces Alzheimer's pathology and rescues neuronal loss *in vivo*. Neurobiol Dis 2009; 35: 359–67.
- **130** Oz M, Lorke DE, Petroianu GA. Methylene blue and Alzheimer's disease. Biochem Pharmacol 2009; 78: 927–32.
- **131** Wischik C, Staff R. Challenges in the conduct of disease-modifying trials in AD: practical experience from a phase 2 trial of Tau-aggregation inhibitor therapy. J Nutr Health Aging 2009; 13: 367–9.
- **132** Cattaneo A, Capsoni S, Paoletti F. Towards non invasive nerve growth factor therapies for Alzheimer's disease. J Alzheimers Dis 2008; 15: 255–83.
- 133 Eriksdotter Jonhagen M, Nordberg A, Amberla K, Backman L, Ebendal T, Meyerson B, Olson L, Seiger A, Shigeta M, Theodorsson E, Viitanen M, Winblad B, Wahlund LO. Intracerebroventricular infusion of nerve growth factor in three patients with Alzheimer's disease. Dement Geriatr Cogn Disord 1998; 9: 246–57.
- **134** Olson L, Nordberg A, von Holst H, Backman L, Ebendal T, Alafuzoff I, Amberla K, Hartvig P, Herlitz A, Lilja A. Nerve

- growth factor affects 11C-nicotine binding, blood flow, EEG, and verbal episodic memory in an Alzheimer patient (case report). J Neural Transm Park Dis Dement Sect 1992; 4: 79–95.
- 135 Tuszynski MH, Thal L, Pay M, Salmon DP, U HS, Bakay R, Patel P, Blesch A, Vahlsing HL, Ho G, Tong G, Potkin SG, Fallon J, Hansen L, Mufson EJ, Kordower JH, Gall C, Conner J. A phase 1 clinical trial of nerve growth factor gene therapy for Alzheimer disease. Nat Med 2005; 11: 551–5.
- **136** Bishop KM, Hofer EK, Mehta A, Ramirez A, Sun L, Tuszynski M, Bartus RT. Therapeutic potential of CERE-110 (AAV2-NGF): targeted, stable, and sustained NGF delivery and trophic activity on rodent basal forebrain cholinergic neurons. Exp Neurol 2008; 211: 574–84.
- 137 Nagahara AH, Bernot T, Moseanko R, Brignolo L, Blesch A, Conner JM, Ramirez A, Gasmi M, Tuszynski MH. Long-term reversal of cholinergic neuronal decline in aged non-human primates by lentiviral NGF gene delivery. Exp Neurol 2009; 215: 153–9.
- **138** Mandel RJ. CERE-110, an adeno-associated virus-based gene delivery vector expressing human nerve growth factor for the treatment of Alzheimer's disease. Curr Opin Mol Ther 2010; 12: 240–7.
- **139** Eriksdotter JM, Linderoth B, Almqvist P, Lind G, Aladellie L, Nordberg A, Kadir A, Jelic V, Seiger A, Wahlberg L. Therapy of Alzheimer's disease with NGF. Neurobiol Aging 2010; 31: (Suppl. 1): S9.
- **140** Reddy PH, Beal MF. Amyloid beta, mitochondrial dysfunction and synaptic damage: implications for cognitive decline in aging and Alzheimer's disease. Trends Mol Med 2008; 14: 45–53.
- **141** Su B, Wang X, Bonda D, Perry G, Smith M, Zhu X. Abnormal mitochondrial dynamics a novel therapeutic target for Alzheimer's disease? Mol Neurobiol 2010; 41:87–96.
- **142** Zhang S, Hedskog L, Petersen CA, Winblad B, Ankarcrona M. Dimebon (latrepirdine) enhances mitochondrial function and protects neuronal cells from death. J Alzheimers Dis 2010; 21: 389–402.
- 143 Doody RS, Gavrilova SI, Sano M, Thomas RG, Aisen PS, Bachurin SO, Seely L, Hung D. Effect of dimebon on cognition, activities of daily living, behaviour, and global function in patients with mild-to-moderate Alzheimer's disease: a randomised, double-blind, placebo-controlled study. Lancet 2008; 372: 207–15.
- **144** Bezprozvanny I. The rise and fall of dimebon. Drug News Perspect 2010; 23: 518–23.
- **145** Bastianetto S, Krantic S, Chabot JG, Quirion R. Possible involvement of programmed cell death pathways in the neuroprotective action of polyphenols. Curr Alzheimer Res 2011; 8: 445–51.
- **146** Schmidt AM, Sahagan B, Nelson RB, Selmer J, Rothlein R, Bell JM. The role of RAGE in amyloid-beta peptide-mediated pathology in Alzheimer's disease. Curr Opin Investig Drugs 2009; 10: 672–80.



- **147** Cho HJ, Son SM, Jin SM, Hong HS, Shin DH, Kim SJ, Huh K, Mook-Jung I. RAGE regulates BACE1 and Abeta generation via NFAT1 activation in Alzheimer's disease animal model. FASEB J 2009; 23: 2639–49.
- **148** Yan SD, Bierhaus A, Nawroth PP, Stern DM. RAGE and Alzheimer's disease: a progression factor for amyloid-beta-induced cellular perturbation? J Alzheimers Dis 2009; 16: 833–43.
- **149** Laxton AW, Tang-Wai DF, McAndrews MP, Zumsteg D, Wennberg R, Keren R, Wherrett J, Naglie G, Hamani C,

- Smith GS, Lozano AM. A phase I trial of deep brain stimulation of memory circuits in Alzheimer's disease. Ann Neurol 2010; 68: 521–34.
- 150 Vellas B, Aisen PS, Sampaio C, Carrillo M, Scheltens P, Scherrer B, Frisoni GB, Weiner M, Schneider L, Gauthier S, Wied CC, Hendrix S, Feldman H, Cedarbaum J, Petersen R, Siemers E, Andrieu S, Prvulovic D, Touchon J, Hampel H. Prevention trials in Alzheimer's disease: an EU-US task force report. Prog Neurobiol 2011; DOI: 10.1016/j.pneurobio. 2011.08.014.